

# DETERMINATION OF PHOTOTOXICITY PROPERTIES OF DIFFERENT COMPOUNDS USING A FULL THICKNESS SKIN MODEL (AST-2000)

J. J. Hoffmann<sup>1</sup>, E. Heisler<sup>2</sup>, P. Peters<sup>1</sup>; S. Karpinski<sup>1</sup>, H.-W. Vohr<sup>2</sup>

<sup>1</sup>CellSystems Biotechnologie Vertrieb GmbH, St. Katharinen, Germany, <sup>2</sup>Bayer Health Care, Institute of Genetic & Molecular Toxicology, Wuppertal, Germany  
Tel.: ++49-(0)-2241-854420 Fax: ++49-(0)-2241-854421 E-Mail: jens.hoffmann@cellsystems.de

## INTRODUCTION

**AST-2000**

- Organotypical coculture
- Human fibroblasts in dermis equivalent
- Human keratinocytes in differentiated epidermis
- Coculture on filter membrane, pore diameter 0.2 µm
- Surface of the skin equivalent: 0.253 inch<sup>2</sup>

**Applications**

- Differentiation studies
- Survey of barrier function
- Irritation studies
- Migration and wound healing studies
- Toxicity / gene-toxicity studies

**Advantages**

- Autonomy from donor material
- Constant conditions
- Reproducible results
- Provides physiological studies
- Similar to the in-vivo-skin

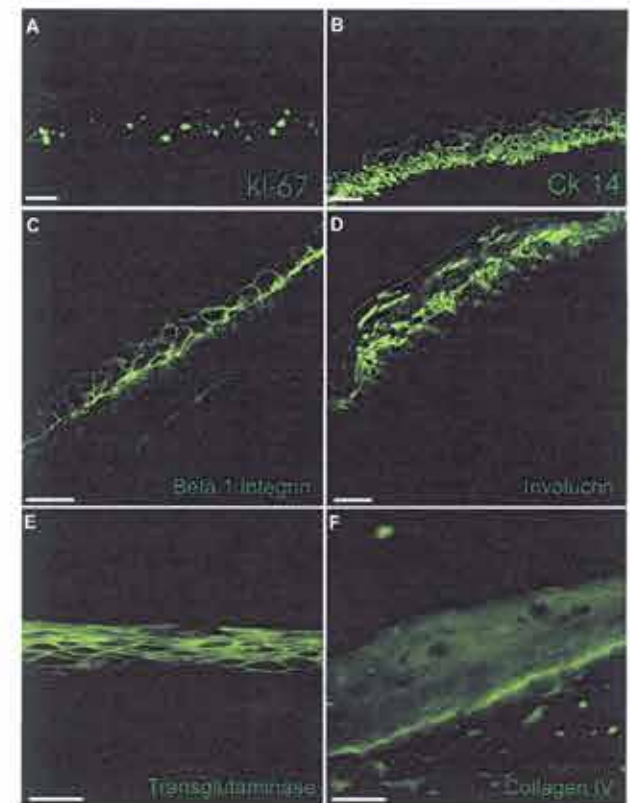
**References:**

Hoffmann, J., Heisler, E., Karpinski, S., Peters, P., Vohr, H.-W. Reconstructed human skin (AST-2000) as a tool for pharmacotoxicology. *ATLA*, 21, 302 (1999)

Hoffmann, J., Vohr, H.-W. Local immune response in vitro: Skin models for the discrimination between irritative and sensitization. *Toxicology*, 254, 209 (1)

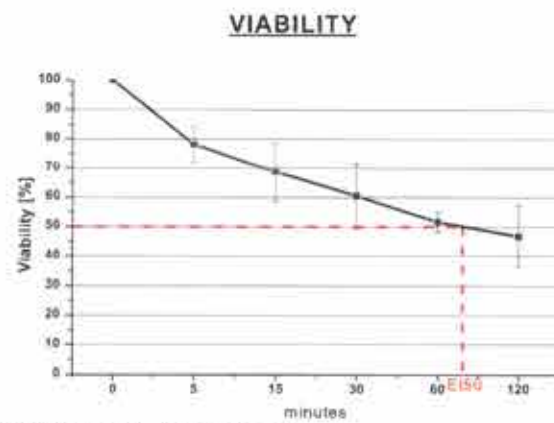
**Fig. 1: AST-2000:** Comparison of native human (A) skin and AST-2000 (B). Paraffin embedding followed by H&E staining. Bar 10 µm. AST-2000 is cultured in inserts on a polycarbonate membran (C). At the end of the culture period the skin equivalent is about 0,63 cm<sup>2</sup> in size.

## EXPRESSION OF SPECIFIC MARKERS

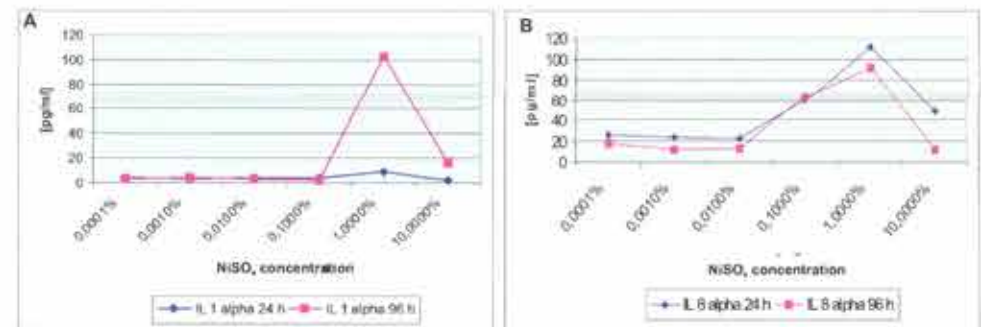


**Fig. 2: Detection of specific markers by immunofluorescence:** Cryosections of AST-2000 treated with antibodies directed against KI-67 (A), cytokeratin 14 (B), integrin β1 (C), involucrin (D), transglutaminase (E) and collagen IV (F). Confocal images; Bar: 20 µm.

## EFFECT OF SDS

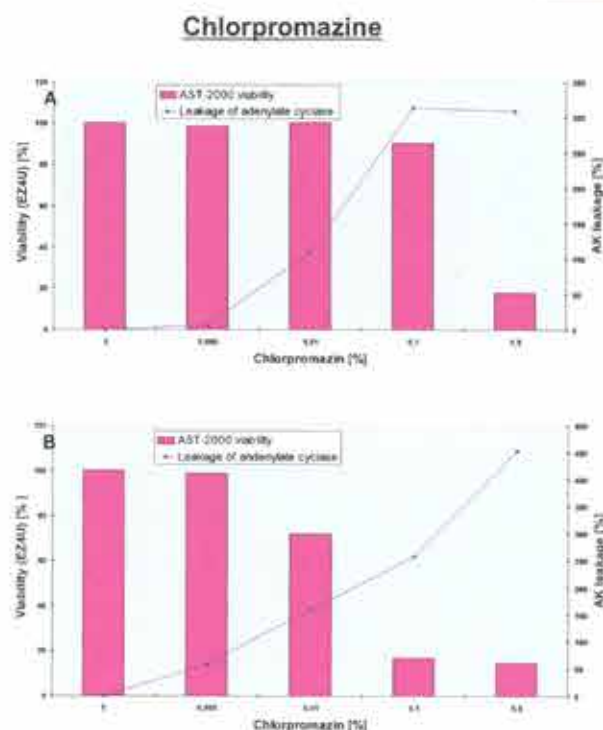


**Fig. 3: Determination of Et50 (1% SDS)**  
5 different lots of AST-2000 were treated for 0; 5; 15; 30; 60 and 120 minutes with 1% SDS (n=5/Lot). Cell viability was determined using a standard MTT-assay leading to an Et50 value (meaning the time after SDS application when cell viability is reduced to 50%) of about 80 minutes.

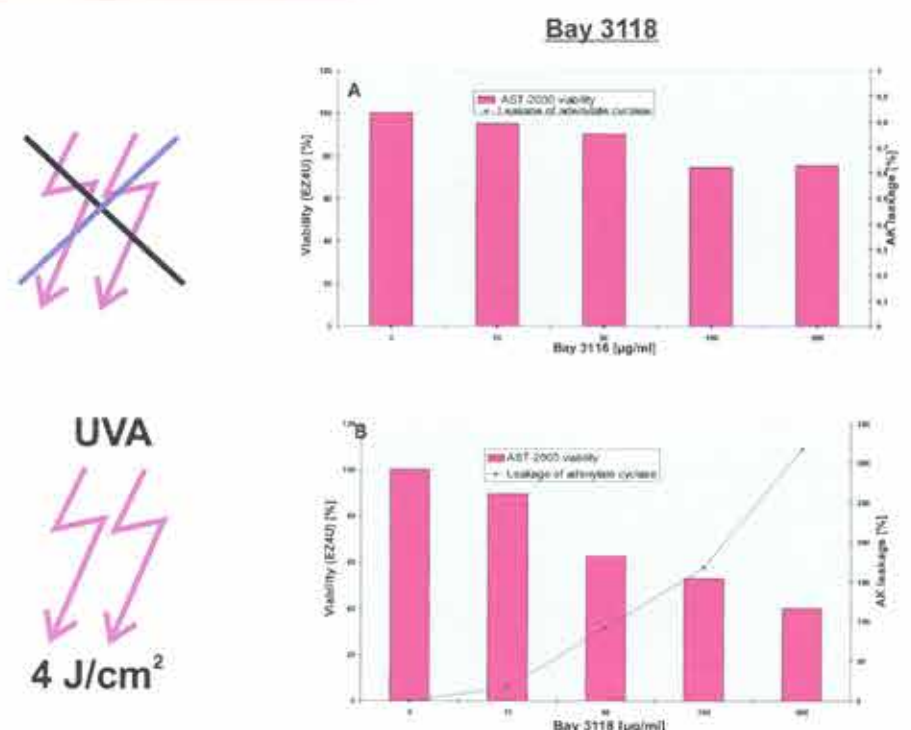


**Fig. 4: Induction of cytokine release:** AST-2000 was treated with NISO, for 24 and 96 hours. The concentration of IL 1 alpha (A) and IL 8 (B) in the culture medium was quantitated by ELISA. Notice that NISO, provoked a specific dose- and time-dependent induction of cell inflammation mediators.

## DETERMINATION OF PHOTOTOXICITY



**Fig. 5: Determination of phototoxic properties of chlorpromazine using AST-2000:** Different concentrations of chlorpromazine were applied topically on the epidermis of AST-2000 followed by UVA radiation (4J/cm<sup>2</sup>) after 20 minutes. After incubation over night the cell viability was determined using the EZ4U Test (Biozol) accompanied by measuring the leakage of adenylate cyclase (AK). Note that cell damage and AK leakage were significant higher in UVA treated AST-2000 as compared to unirradiated control inserts (A).



**Fig. 6: Determination of phototoxic properties of Bay 3118 using AST-2000:** AST-2000 was cultured under normal conditions with medium supplemented with different concentrations of Bay 3118 for 4 hours followed by UVA radiation (4J/cm<sup>2</sup>). After incubation for additional 20 hours the cell viability was determined using the EZ4U Test (Biozol) accompanied by measuring the leakage of adenylate cyclase (AK). Note that cell damage and AK leakage were significant higher in UVA treated AST-2000 as compared to unirradiated control inserts (A).